

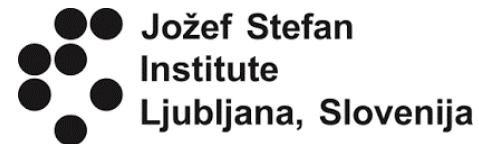
Preparation of tissues for μ XRF/ μ XAS analyses

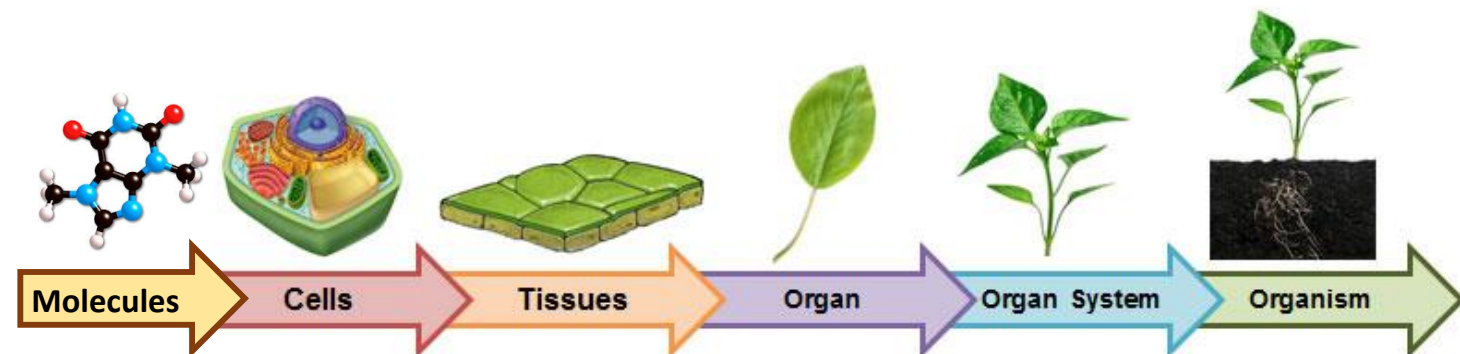
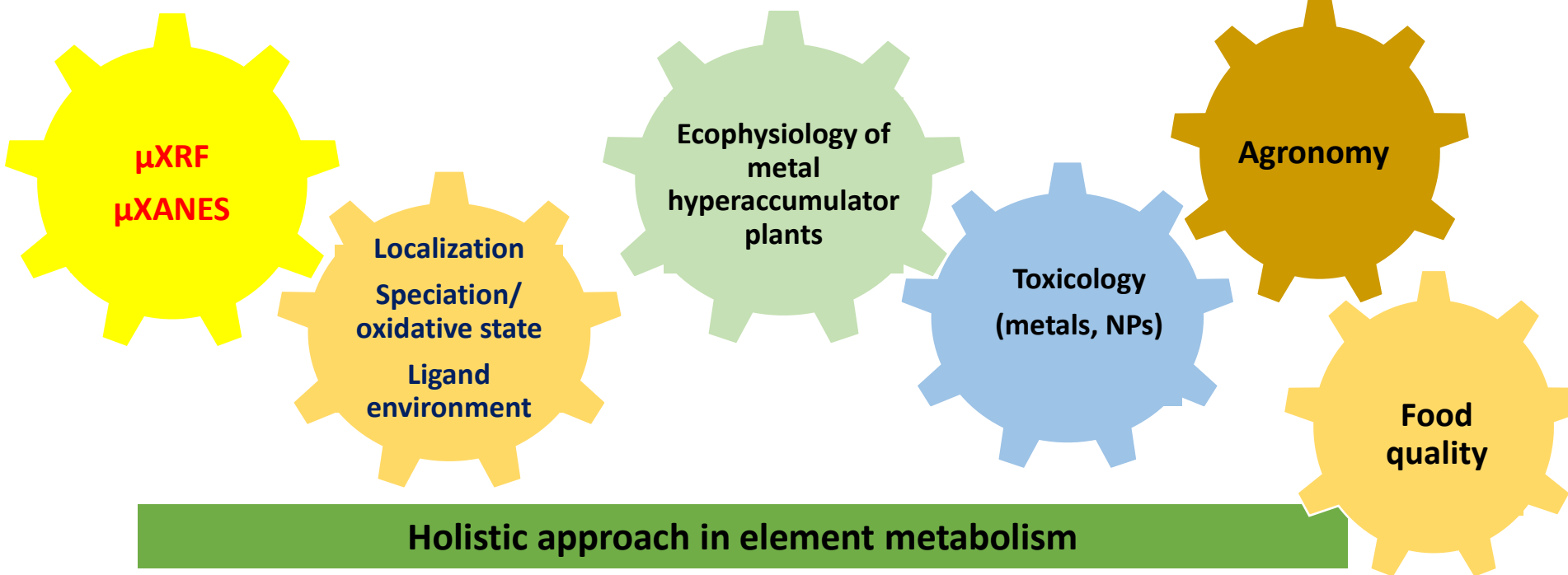
Katarina Vogel-Mikuš

University of Ljubljana, Biotechnical Faculty, Department of Biology

Jozef Stefan institute

* katarina.vogelmikus@bf.uni-lj.si



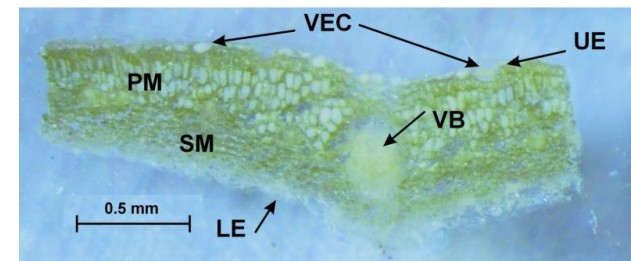


Sample preparation for imaging & μ -XAS



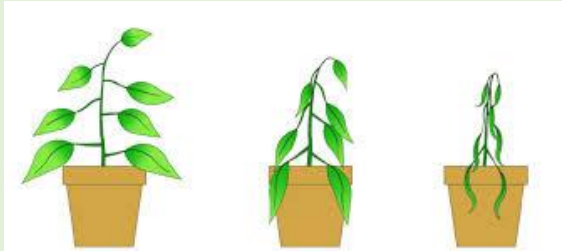
- The main goals to be achieved during sample preparation:
 - preserve local distribution of elements in tissues
 - preserve sample morphology and anatomy
 - preserve metal ligand environment as similar as “in vivo” stage
 - Vacuum compatible samples (for LE-XRF, μ PIXE)

Changes in tissue morphology and chemical redistribution must be limited to dimensions that are smaller than the resolution of the microprobe.



Sampling

- **Good/normal physiological state of an organism** (depending on the experimental conditions)



Case study 1: Changes in elementome during water deprivations



Case study 1: Accumulation of Cd in metal hyperaccumulating plant

- **Short time between organ excision and fixation (activation of degradation enzymes** - proteases, cellulases, pectinases, catechol oxidases)

Sampling

- **Laboratory experiments**

- keeping plants in good conditions
- prevent wilting



- **Field experiments**

- small plants – excavation together with the roots
- Bigger plants (trees) fixation in the field



Sample preparation

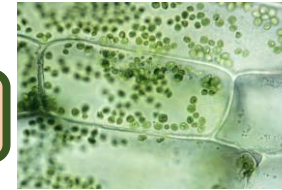


Sample

Organ level

Tissue level

Cell level



Direct measurements

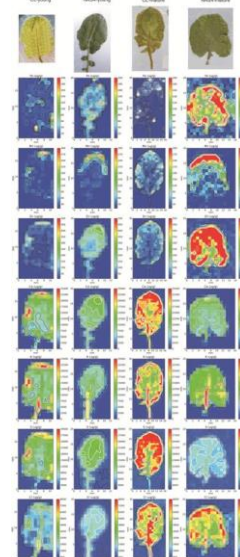
Tissue & cell exposure

Fixation

- „in air“ HE- μ XRF
- Fast acquisition system (MAIA detector), dwell time >10 ms/px

PROBLEMS

- Radiation damage (water, ROS)
- Metabolic changes
- XRF signal coming from different depths
- Combined information across different depths/ physiologically distinct tissues



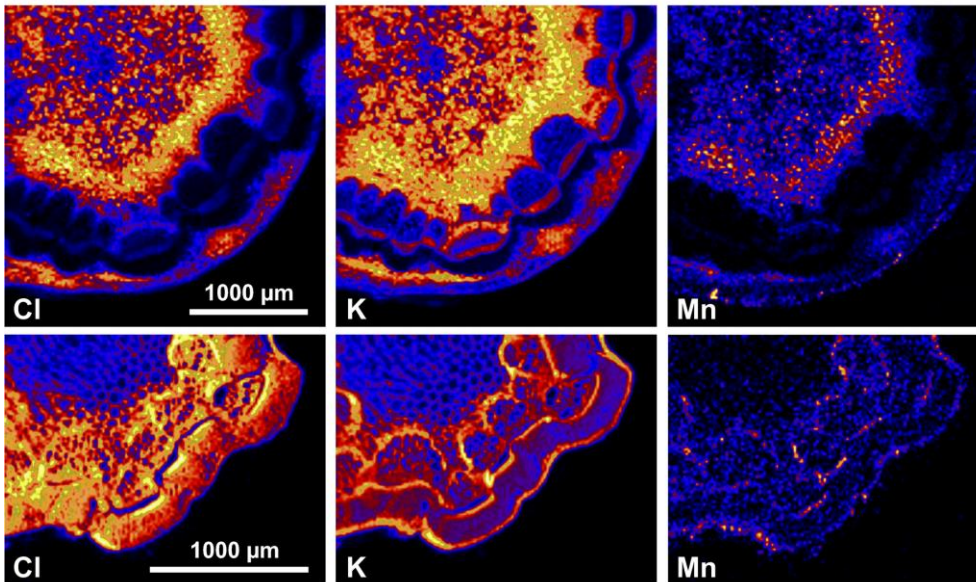
- Vacuum compatibility (μ LE-XRF, μ PIXE)

Ecophysiology - analysis of herbarium specimens

- Air-drying of plant material **NOT SUITABLE!**



(d) Freeze-dried (below)



(e) Air-dried (herbarium, above)

- Distorted morphology & element localization



New Phytologist

Tansley review | Free Access

X-ray elemental mapping techniques for elucidating the ecophysiology of hyperaccumulator plants

Antony van der Ent, Wojciech J. Przybyłowicz, Martin D. de Jonge, Hugh H. Harris, Chris G. Ryan, Grzegorz Tylko, David J. Paterson, Alban D. Barnabas, Peter M. Kopitke, Jolanta Mesjasz-Przybyłowicz

First published: 10 October 2017 | <https://doi.org/10.1111/nph.14810> | Cited by: 10

Fixation

Chemical fixation (TEM)

Fixatives

- PFA
- Ethanol
- methanol

dehydration

embedding

Sectioning (<20 um)
Ultrathin cuttings

Cryofixation

Freezing

Sectioning
(Cryostat)

Direct measurements
Frozen-hydrated state

lyophilization



Comparison between chemical and cryofixation



LA-ICPMS

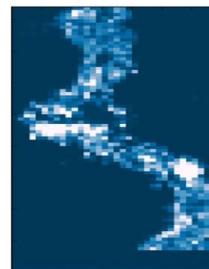
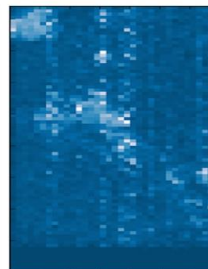
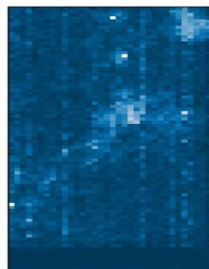
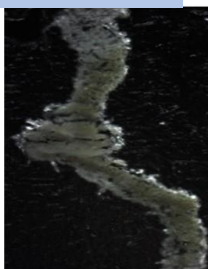
Ca

K

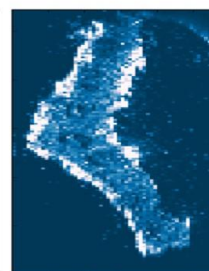
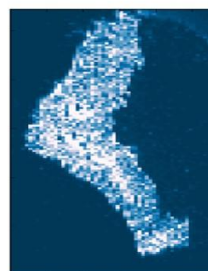
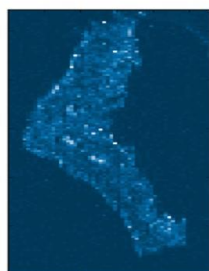
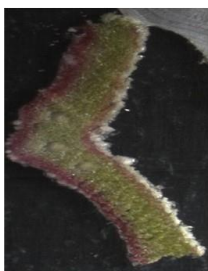
P

Zn

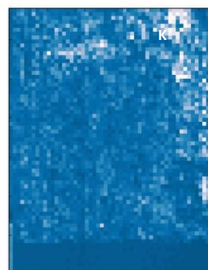
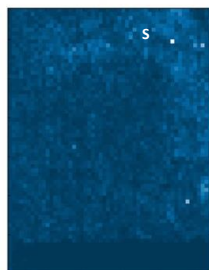
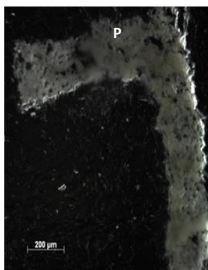
PFA_Zn



Lio_Zn



MeOH_Zn



min

concentration

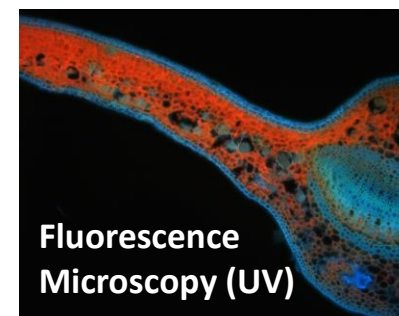
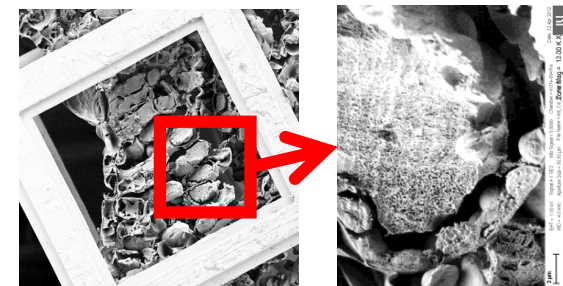
max

Chemical fixation:

- Element redistribution
- Contamination

Cryofixation

- Preservation of structure and function



Fluorescence
Microscopy (UV)

Fixation

Chemical fixation
(Traditional)

Cryofixation

Freezing

Sectioning
(Cryostat)

Fixatives

- PFA
- Ethanol
- methanol

dehydration

embedding

Sectioning (<20 um)
Ultrathin cuttings

Direct measurements
Frozen-hydrated state

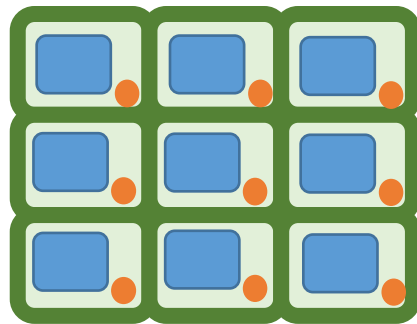
Liophylization
(Freeze-drying)

PROBLEMS

- ice-crystal formation, tissue and cell damage
- In FH – water absorbs LE-XRF, dilution (10x)
- In FD - shrinking during freeze-drying
- In FD Water deprivation may alter ligand environment

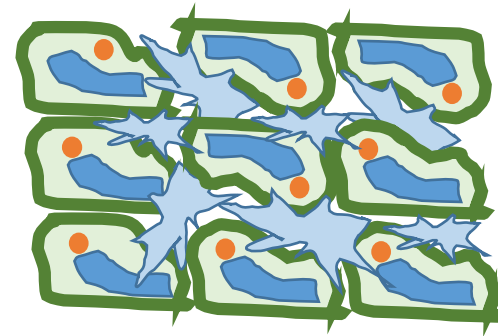
Cryofixation

- Freezing



Environmental conditions

freezing



Ice crystal formation and tissue disintegration

- Prevent ice crystal formation and tissue and cell damage – increase freezing speed - vitrification
- Freezing in LN2 is not suitable

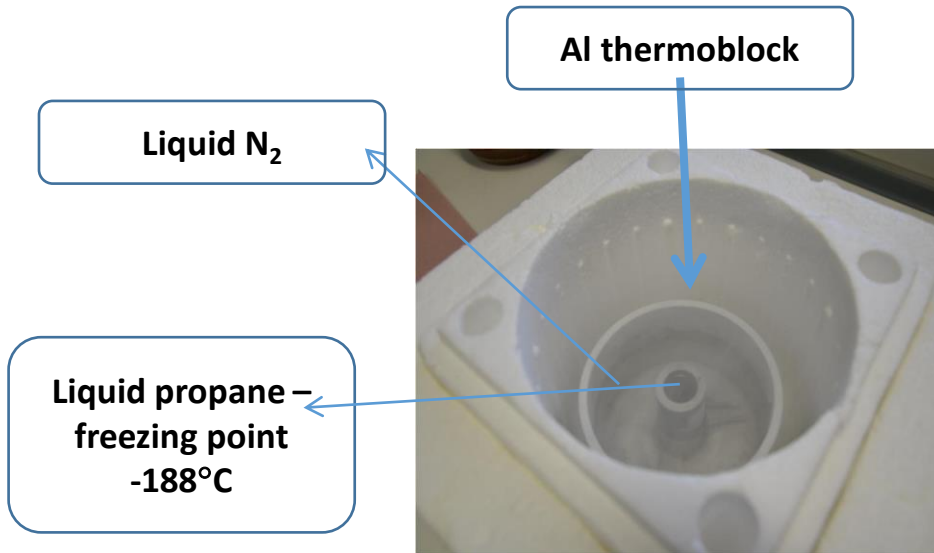


Freezing

- Plunge freezing - cryogenes
- Metal mirror freezing (slam freezing)
- High pressure freezing

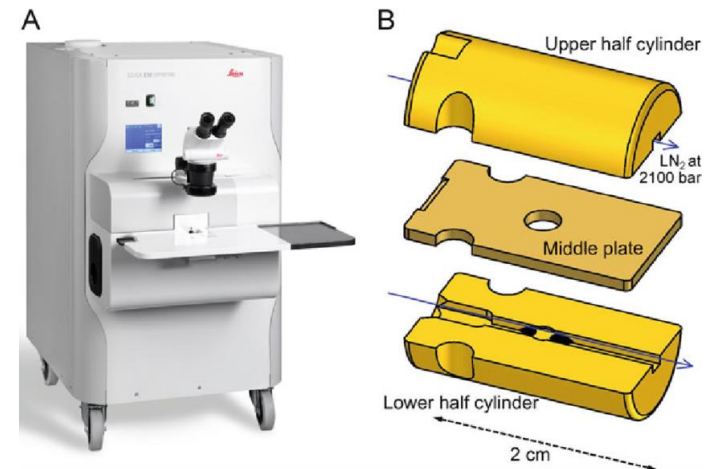


Metal mirror freezing



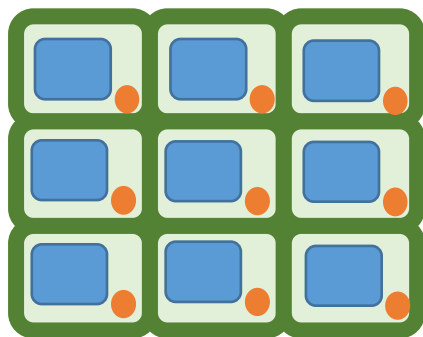
Also ethane, isopentane

Plunge freezing

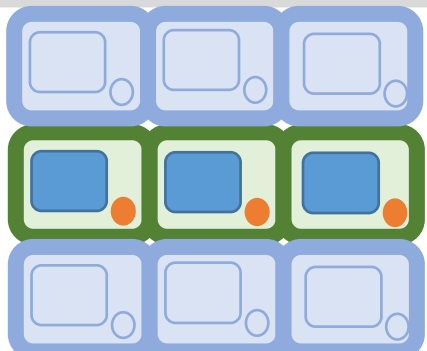


High pressure freezing

Metal mirror freezing



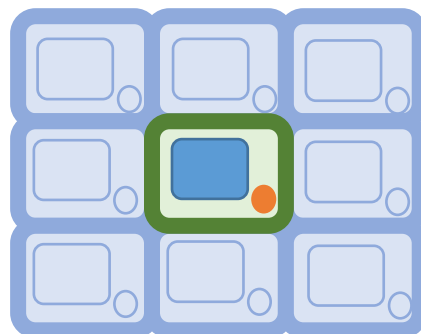
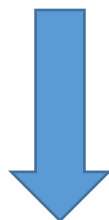
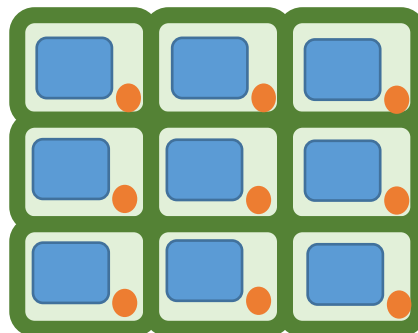
Metal block cooled by LN₂



Metal block cooled by LN₂

Zone of vitrification cca. 10-20 μm

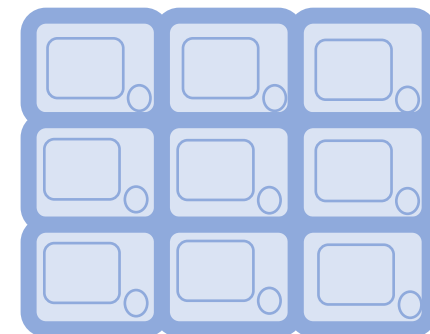
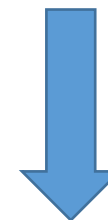
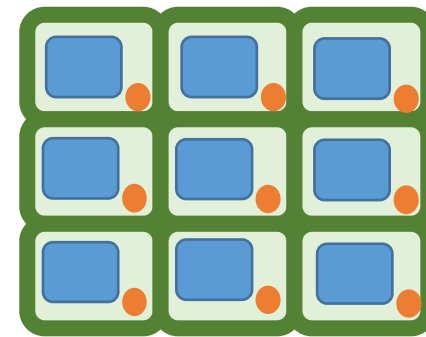
Plunge freezing in propane



Vitrification of up to 100 μm

Efficient only with very small pieces (<100 μm)

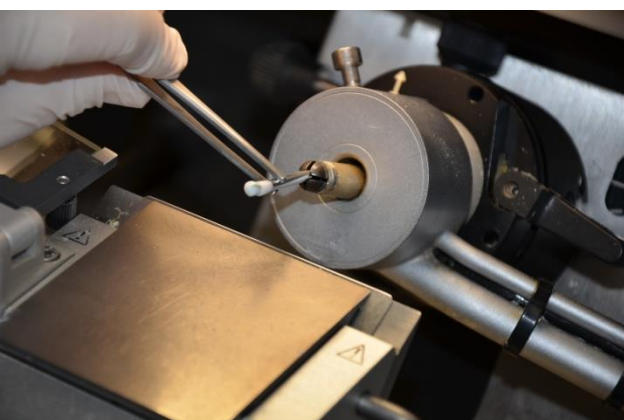
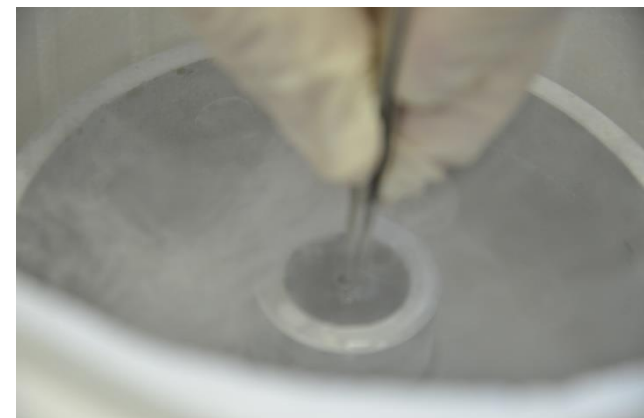
High pressure freezing (NO pre-treatment)



Vitrification of up to 600 μm

Micro-PIXE elemental mapping for ionome studies of crop plants

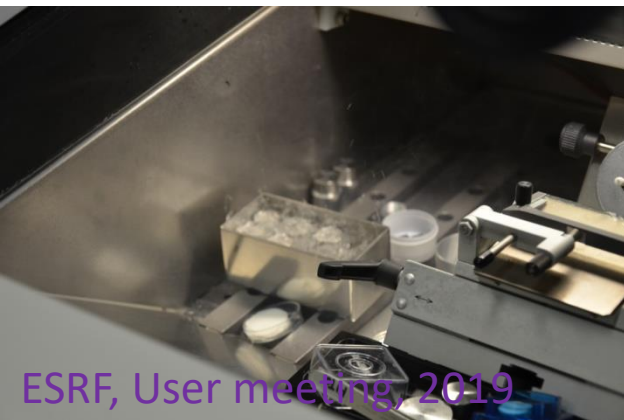
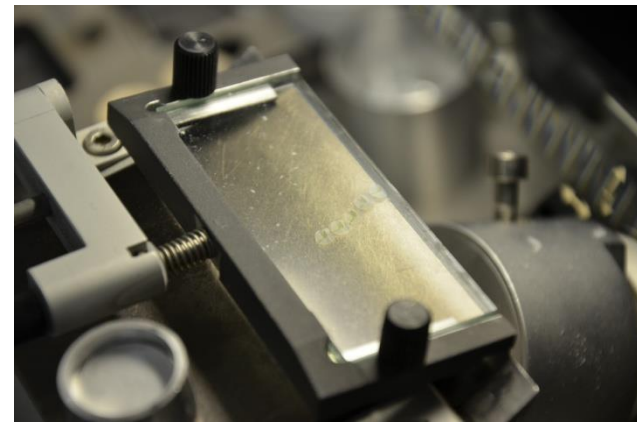
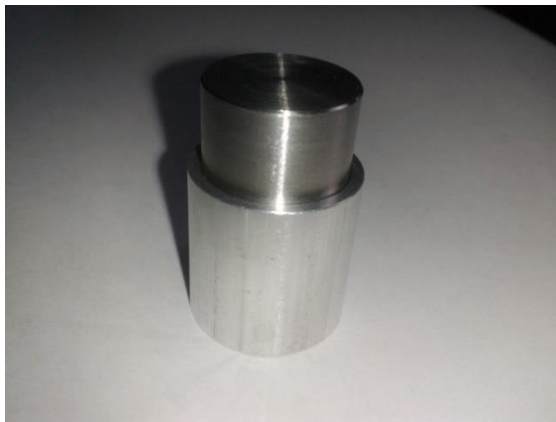
Katarina Vogel-Mikuš, Paula Pongrac and Primož Pelicon



Cryo
Freeze-dried

Freeze-drying

- Should be performed gradually from -196°C to 25°C and slowly to prevent shrinking of the specimens
- Can alter ligand environment (μXAS)



Freeze-drying

- Should be performed **gradually and slowly** (few days) from -196°C to 25°C to prevent shrinking of the specimens
- Computer assisted
- Improved 😊



3rd day – transfer the samples to the highest position – adjustment to room temperature, 24 h

2nd day - transfer the samples to the higher position, 24 h

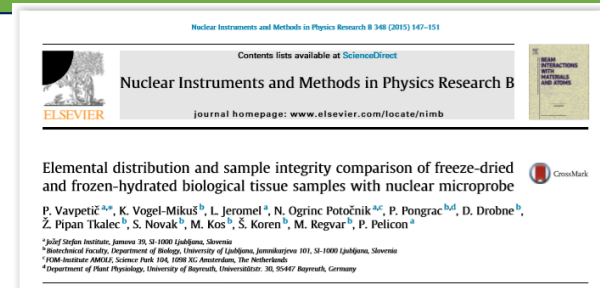
1st day – pour LN2 into the lowest shelf to cool it down, put in the box with samples, leave for 24 h

Shelf temperature

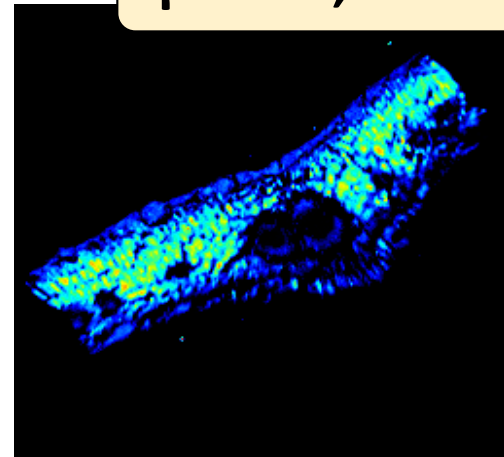
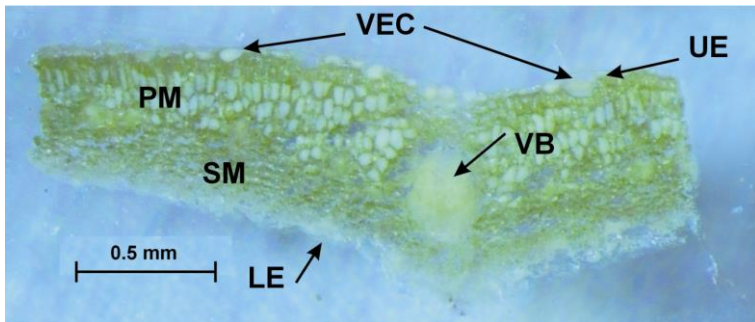
Results – PIXE – tissue/ cell level

N. praecox from natural environment

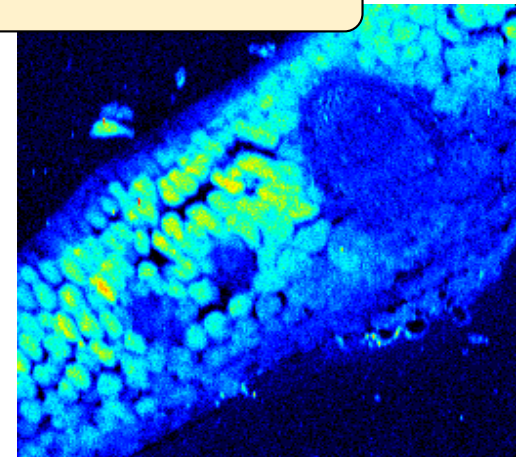
- Well retained morphology and element distribution



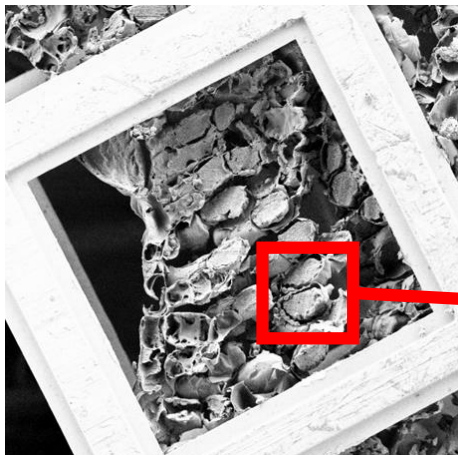
μ-PIXE; leaf cross-sections



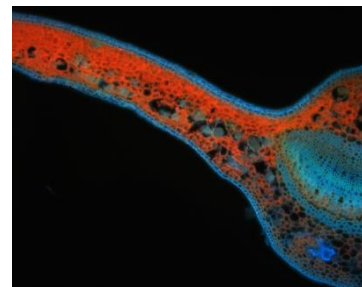
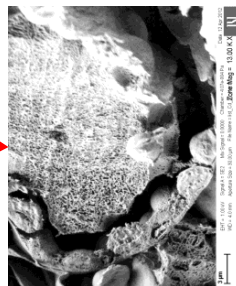
Ca – freeze-dried



Ca – frozen hydrated



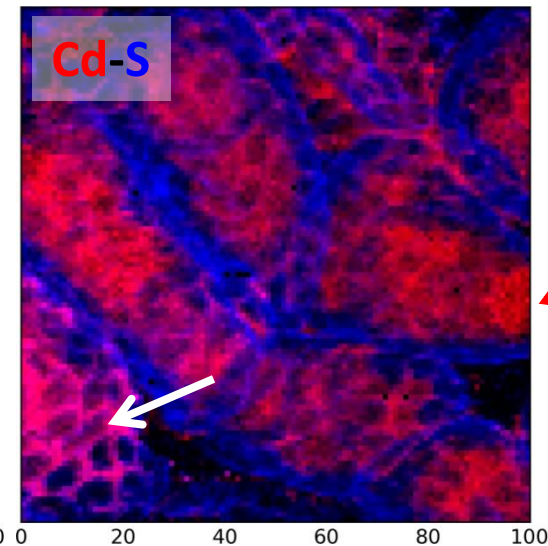
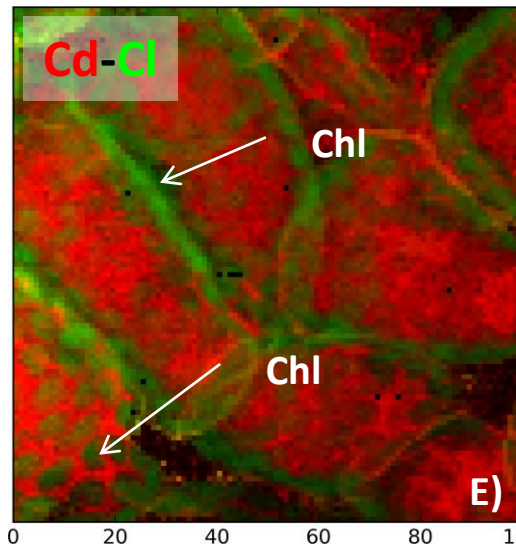
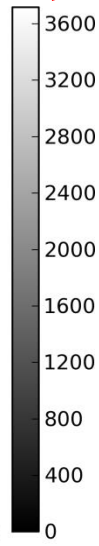
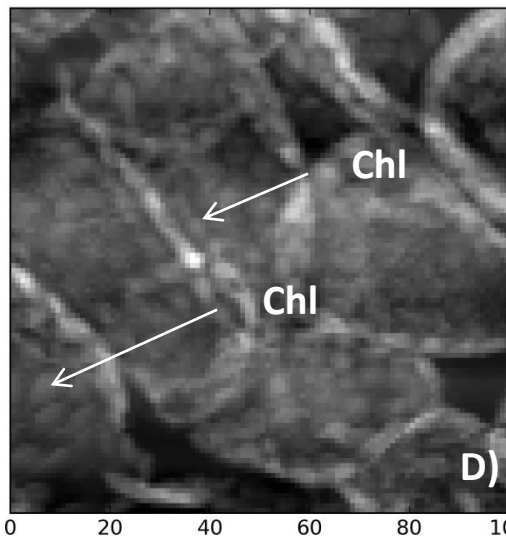
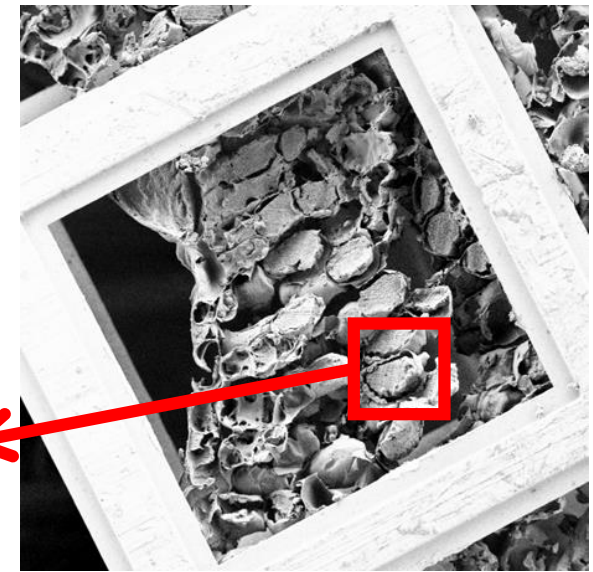
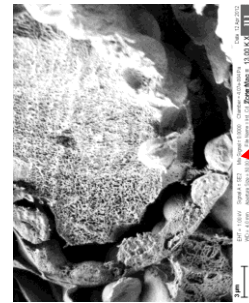
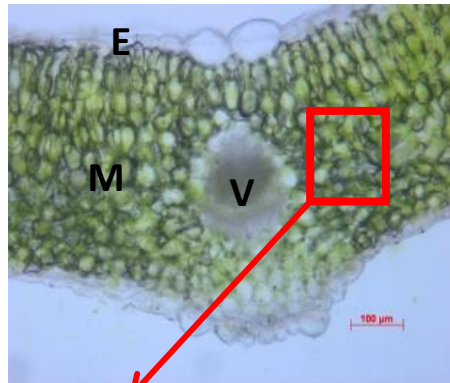
SEM



Fluorescence
Microscopy (UV)

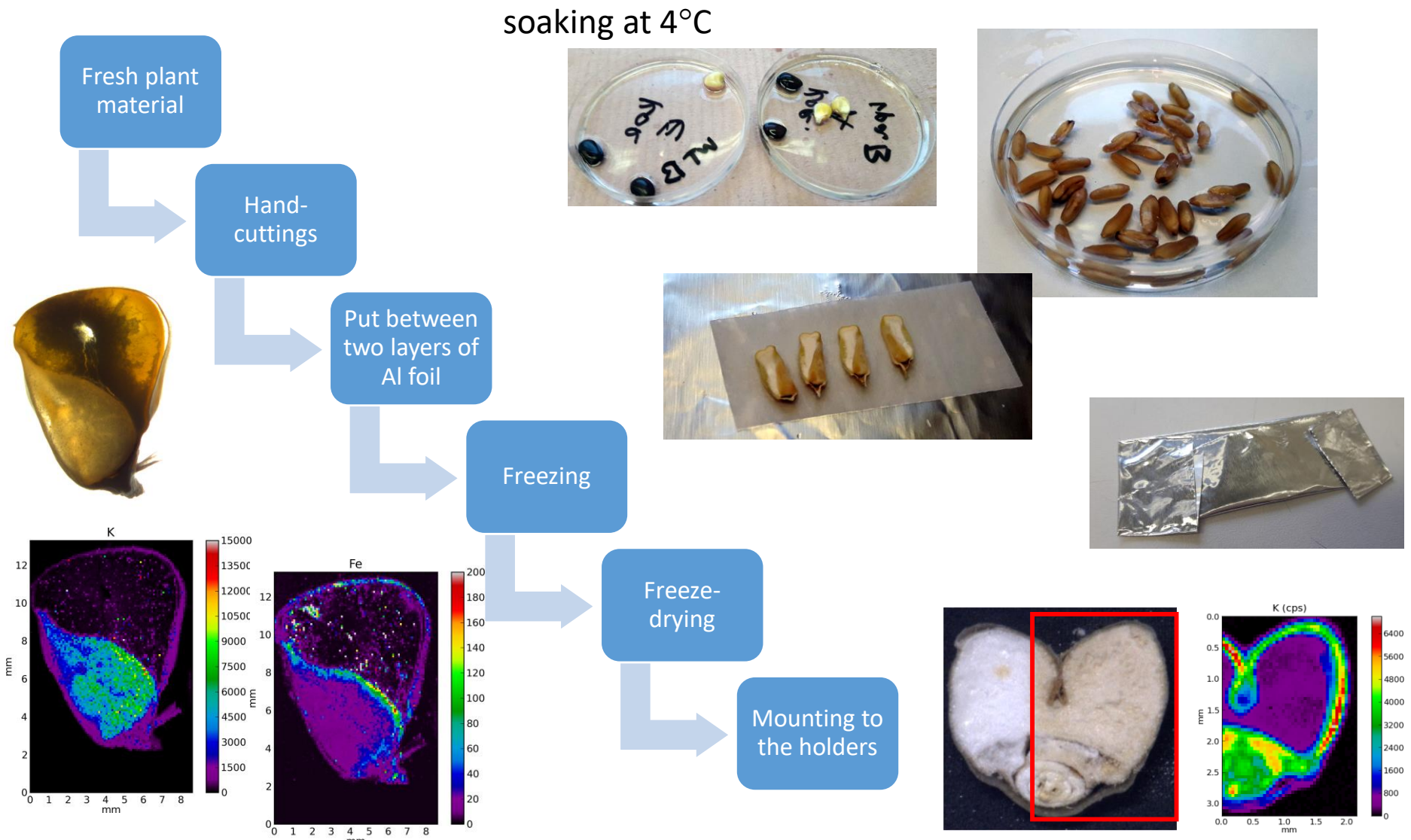
SR- μ XRF - the cell level

T. praecox in hydroponics, Cd



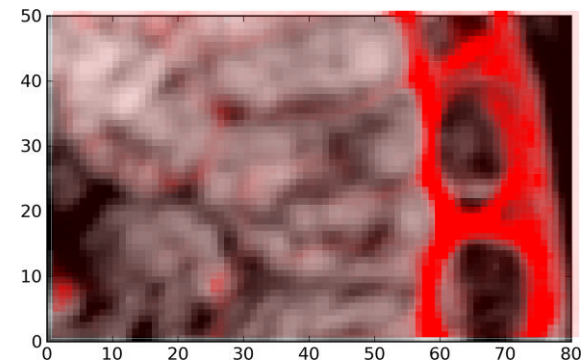
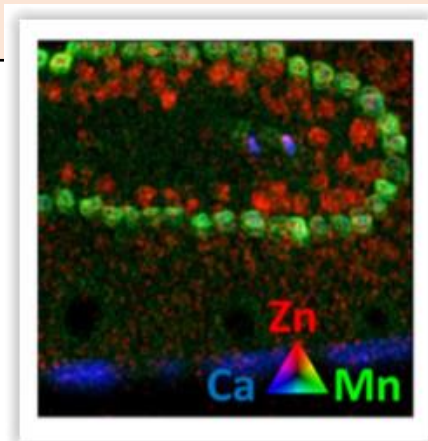
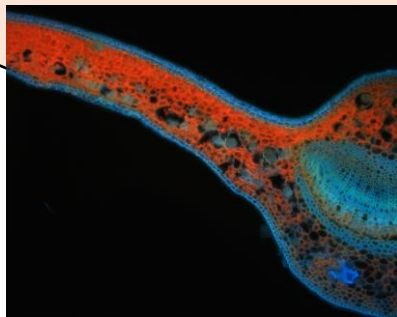
(μ XRF, $E=3.55$ keV, 0.3×0.7 μ m beam), ID 21, ESRF

Hand cutting – organ and tissue levels (seeds)



Conclusions

- Cryofixation is the most suited for μ XRF/ μ XAS
- LN₂ is not suitable cryogen
- Measurements in frozen hydrated state better resemble *in vivo* state, especially for μ -XAS
- Herbarium, air-dried and chemically fixed specimens are not suitable for μ XRF/ μ XAS



Acknowledgements



IAEA
International Atomic Energy Agency



Coworkers:

- ✓ Paula Pongrac (JSI)
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- ✓ Marjana Regvar (BF)
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- ✓ Johannes Teun van Elteren (NIC)
- ✓ Primož Vavpetič (JSI)
- ✓ Mitja Kelemen (JSI)
- ✓ Hiram Castillo & team (ESRF)
- ✓ Alessandra Gianoncelli (ELETTRA)
- ✓ Lorella Pascolo (BGPI)
- ✓

